



**From Concept to Process:
E. coli Metabolic Engineering for Production of
Glucosamine and *N*-Acetylglucosamine**

Bio-Technical Resources (BTR)

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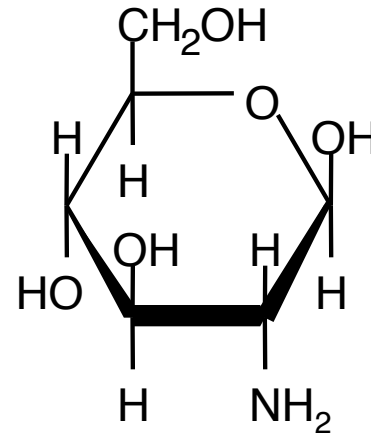
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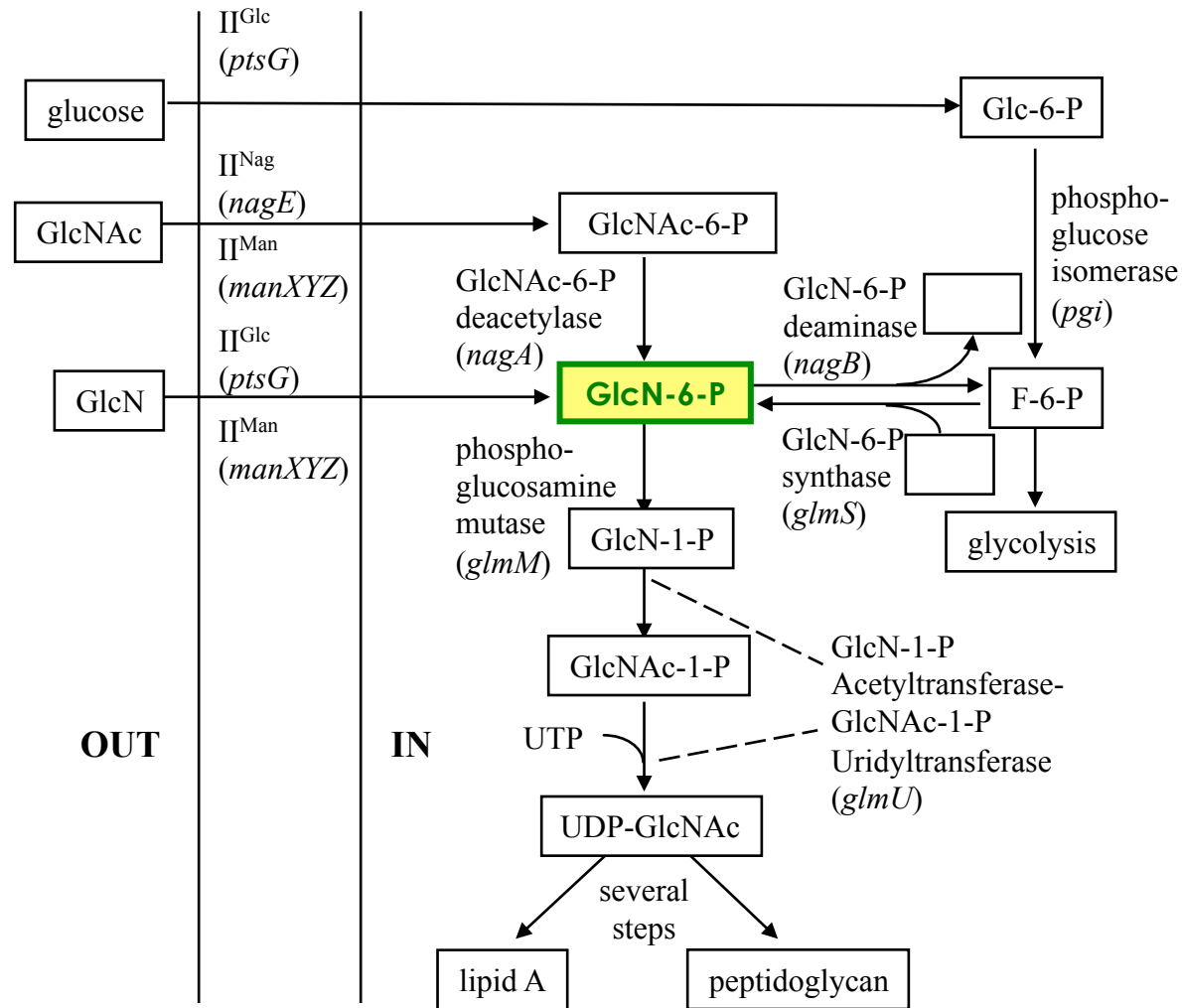
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Glucosamine

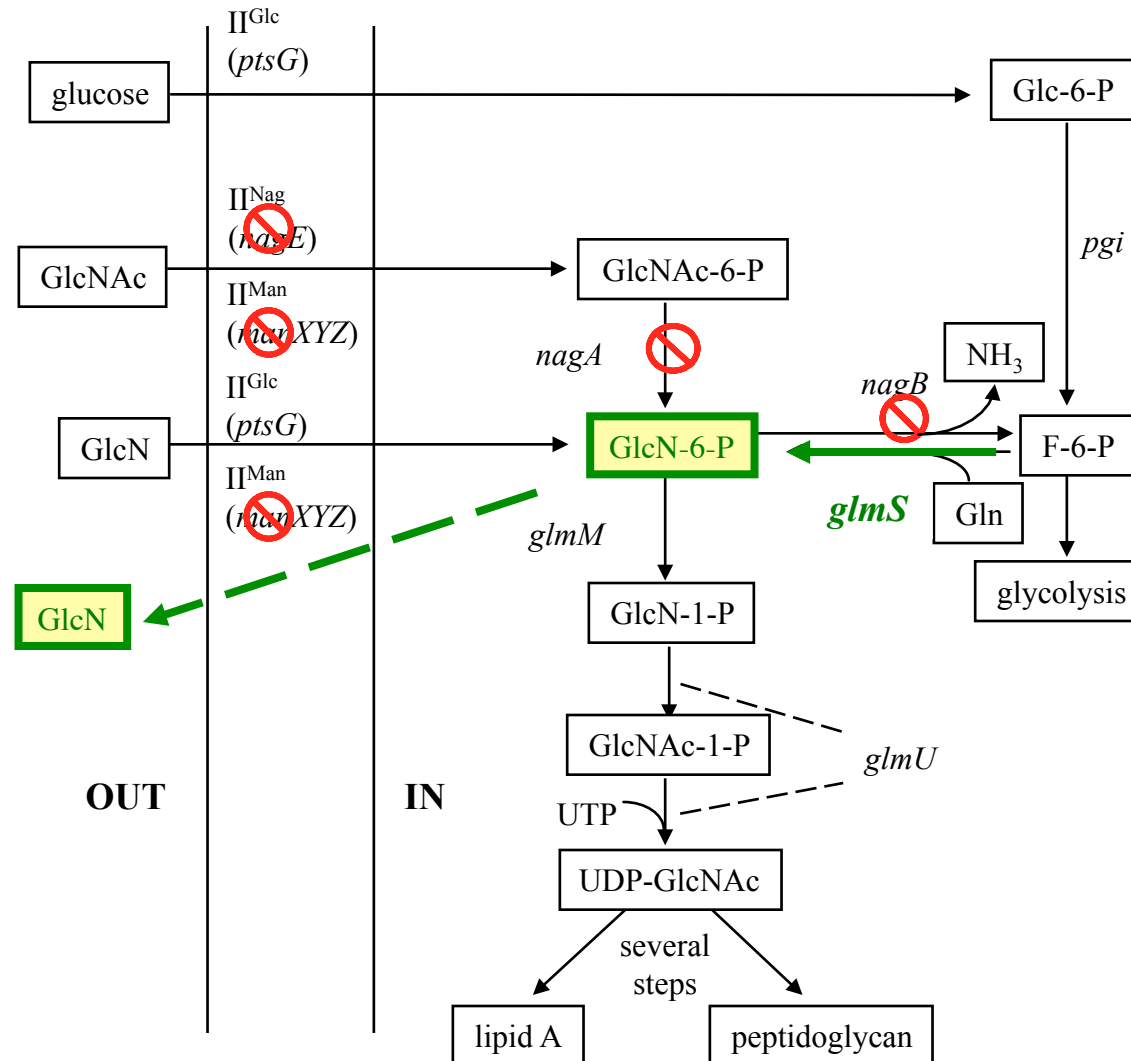


- ❖ Dietary supplement for the treatment of arthritis
- ❖ Increasing market: >\$350 MM/year in the US
- ❖ Current manufacture by hydrolysis of chitin from shellfish
- ❖ Raw material supply limitation and shellfish allergy problem
- ❖ Fermentation process developed by Bio-Technical Resources

Glucosamine Pathway in *E. coli*



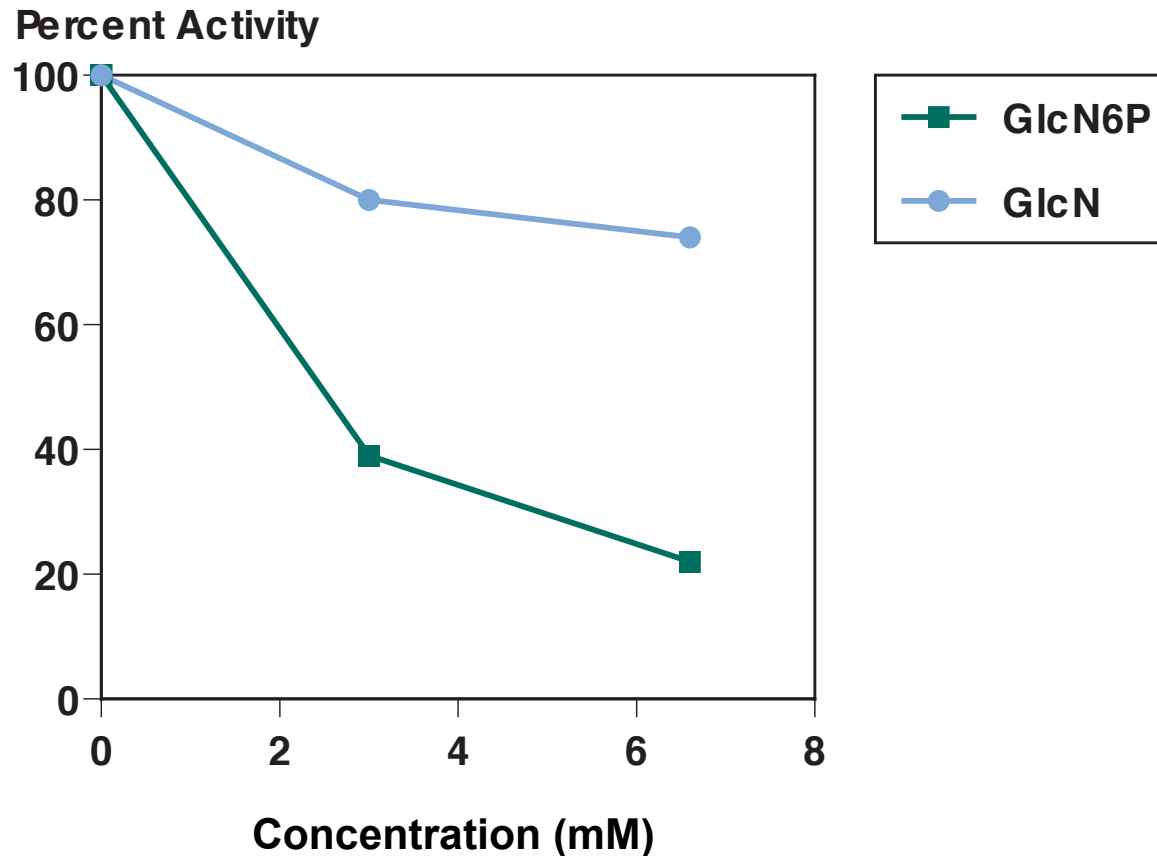
Metabolic Engineering Strategy to Produce Glucosamine



Production of Glucosamine in *E. coli*

<u>Genotype</u>	<u>Glucosamine, mg/L</u>
<u>Shake Flask:</u>	
Host strain ($\Delta nag\ manXYZ$ DE3)	4
T7-glmS plasmid	37
T7-glmS:: $\Delta lacZ$ (integrant)	75
<u>Fermentor:</u>	
T7- glmS:: $\Delta lacZ$ (integrant)	400

Inhibition of GlcN6P Synthase



- ❖ The biosynthesis enzyme GlcN6P Synthase (GlmS) is product-inhibited
- ❖ Activity is lowered to ~20% at 6 mM GlcN6P
- ❖ Directed Evolution was undertaken to generate and select for variants resistant to product inhibition



Generation of Mutant *glmS* Alleles

Error-prone PCR to generate mutations in the
E. coli glmS gene



Library of expression/integration
plasmids in the production host

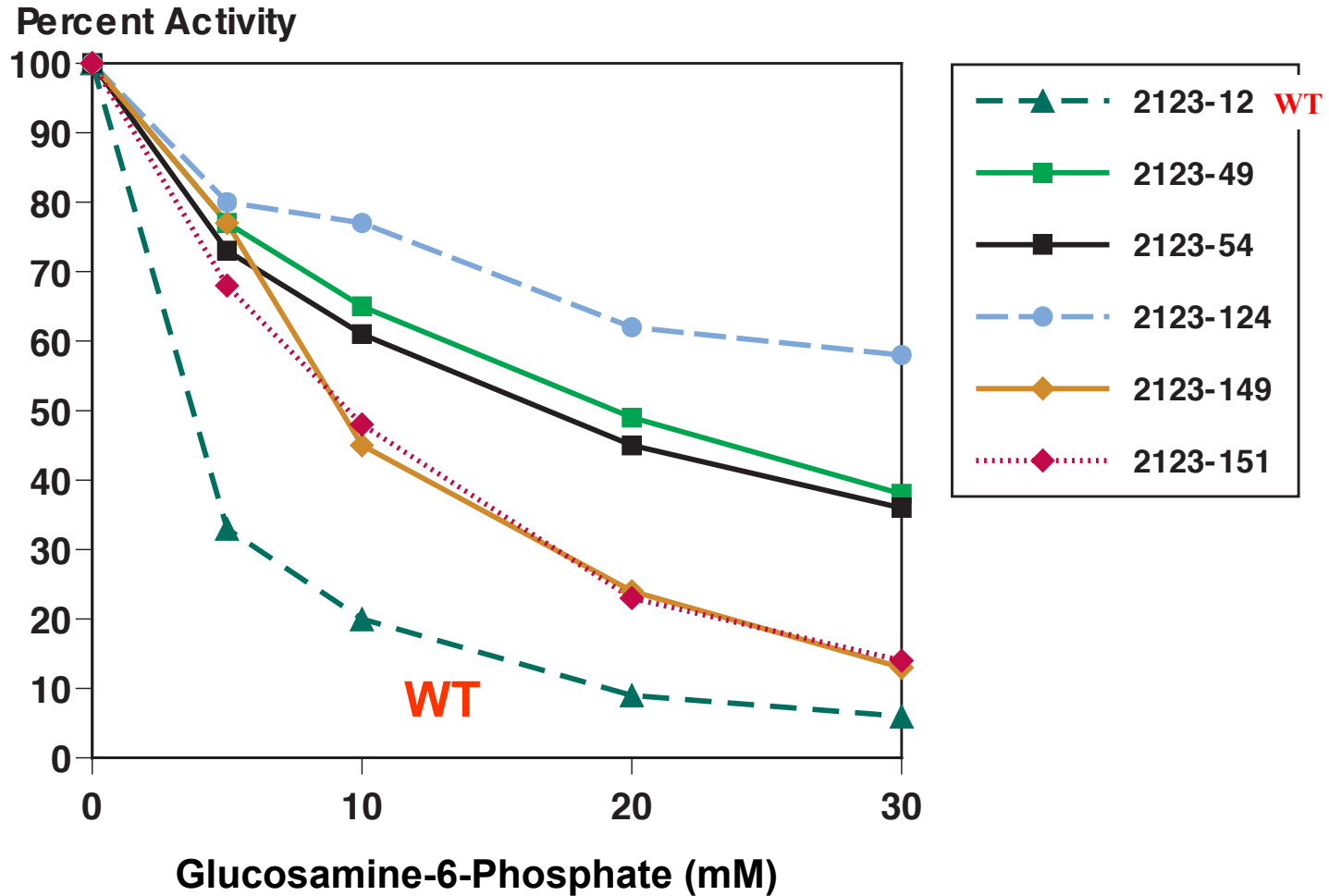


Screening for improved enzymes by
cross-feeding plate assay



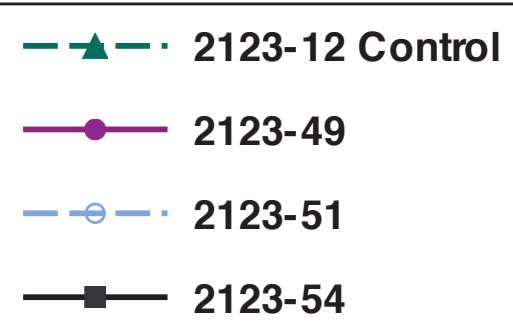
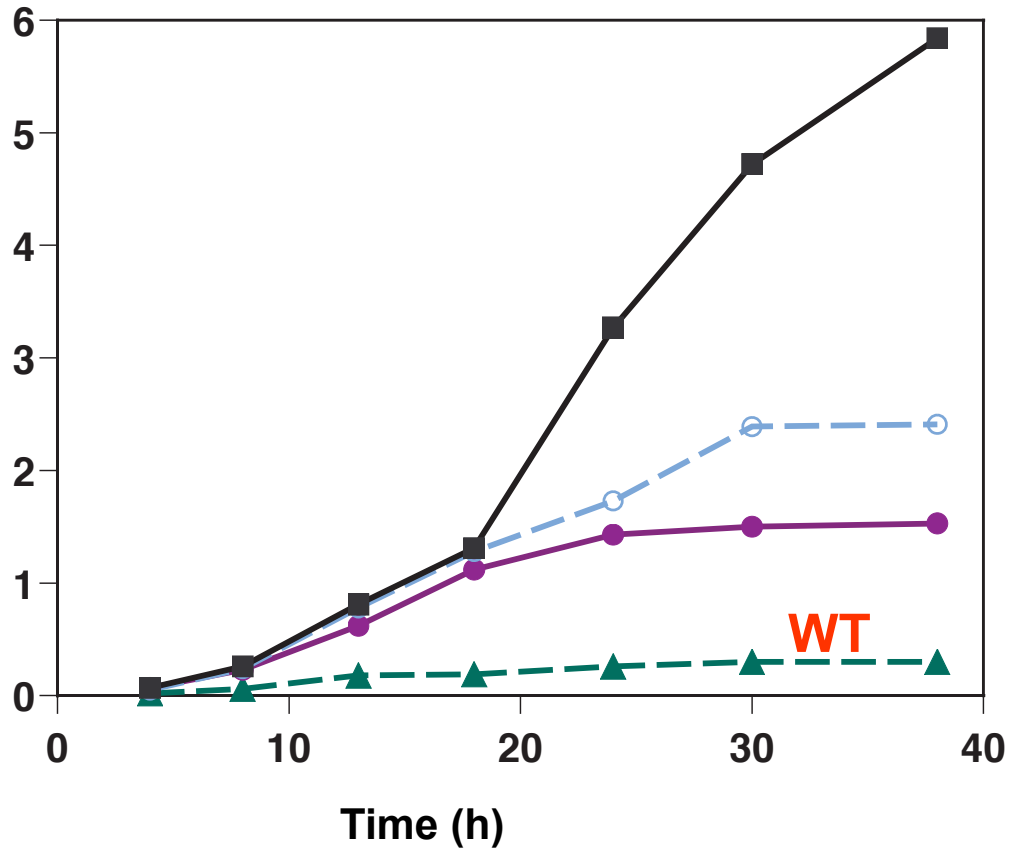
Integration of the mutant *glmS* genes

Product Inhibition of the WT GlmS and Evolved Variants



Glucosamine Production Using End Product-Resistant Mutants

Glucosamine (g/L)



Evolved enzymes
lead to greatly
improved production

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Properties of GlcN6P Synthase in Mutant Strains

<u>Strain</u>	<u>Mutations</u>	<u>Glucosamine Production¹</u>	<u>Specific Activity</u>	<u>Product Resistance²</u>	<u>Thermal Stability³</u>
2123-12	none	0.3	0.39	20	95
2123-49	I4T I272T S450P	4.6	0.38	65	8
2123-54	A39T R250C G472S	7.2	0.42	61	4
2123-124	L469P	5.3	0.008	77	101
2123-149	G472S	0.6	0.49	45	78

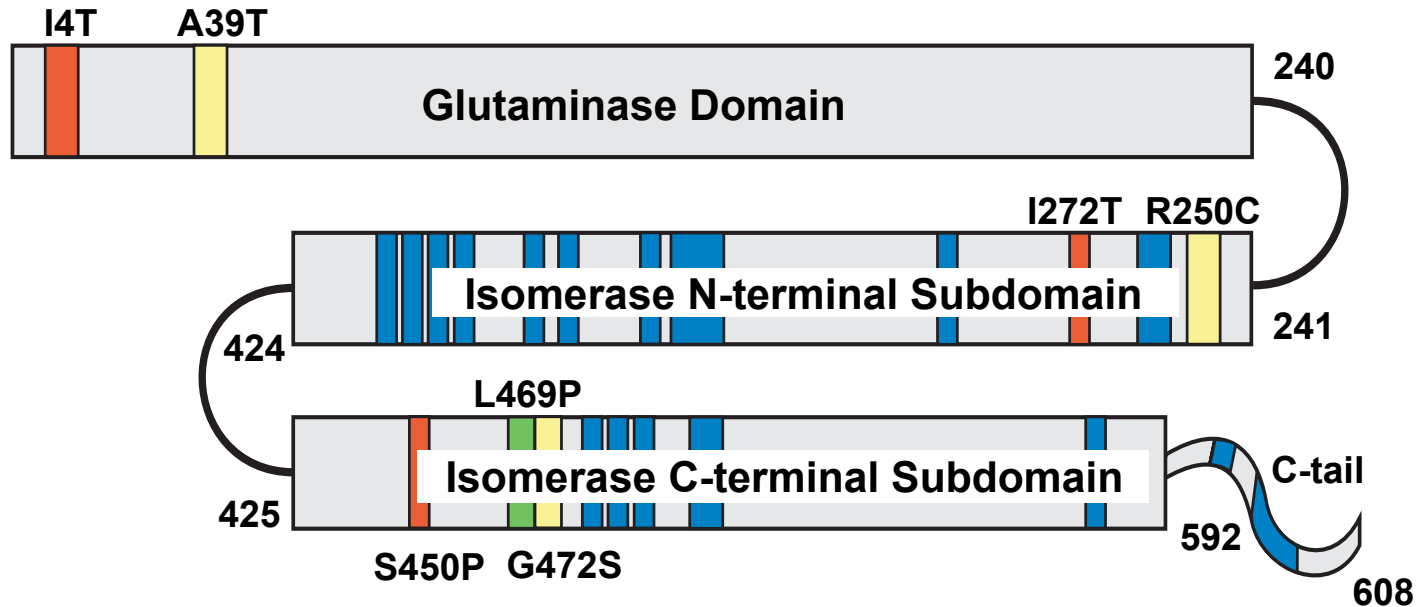
¹Glucosamine production: g/L

²Percent activity with 10 mM glucosamine-6-phosphate

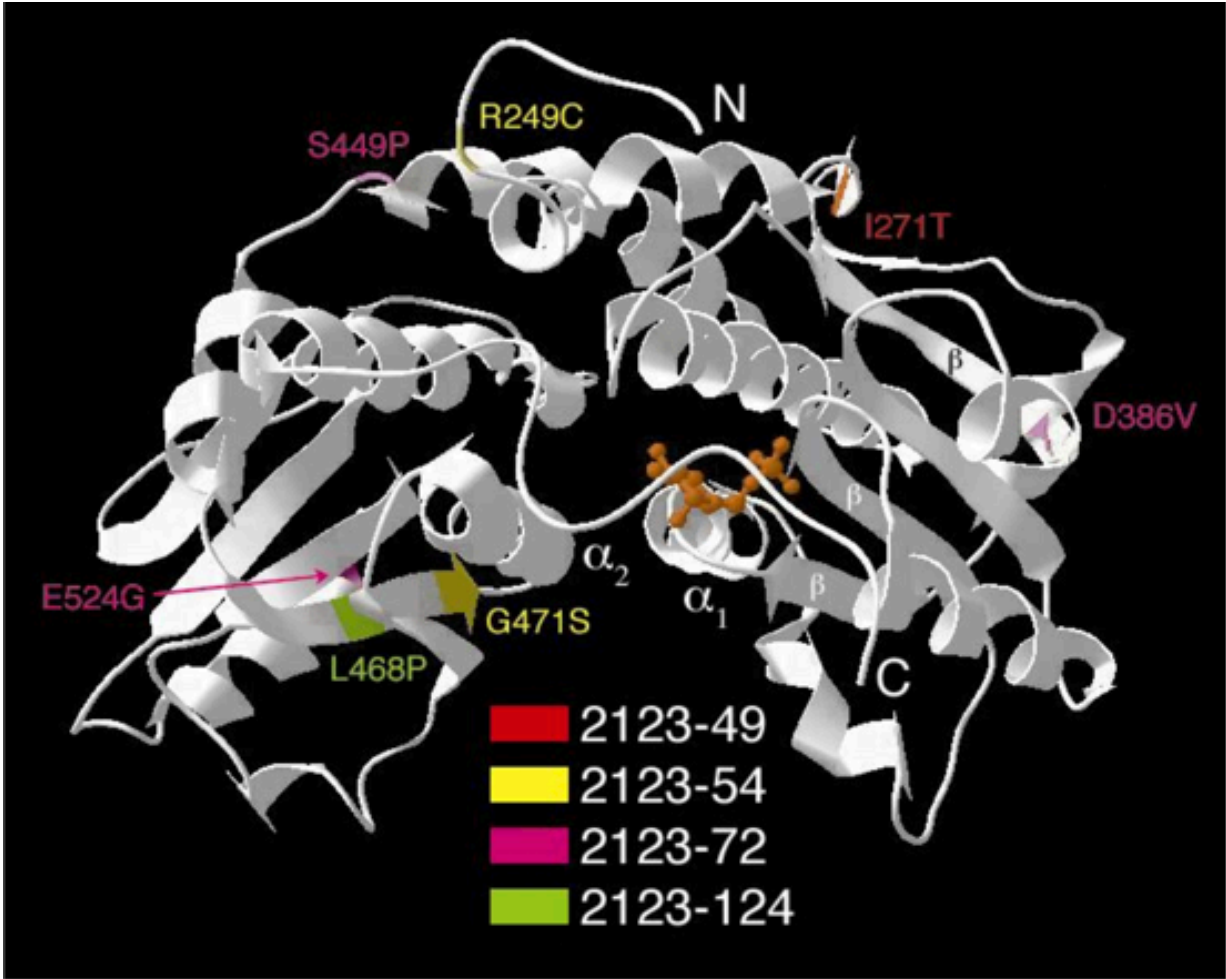
³Percent activity after 30 minutes at 50°C



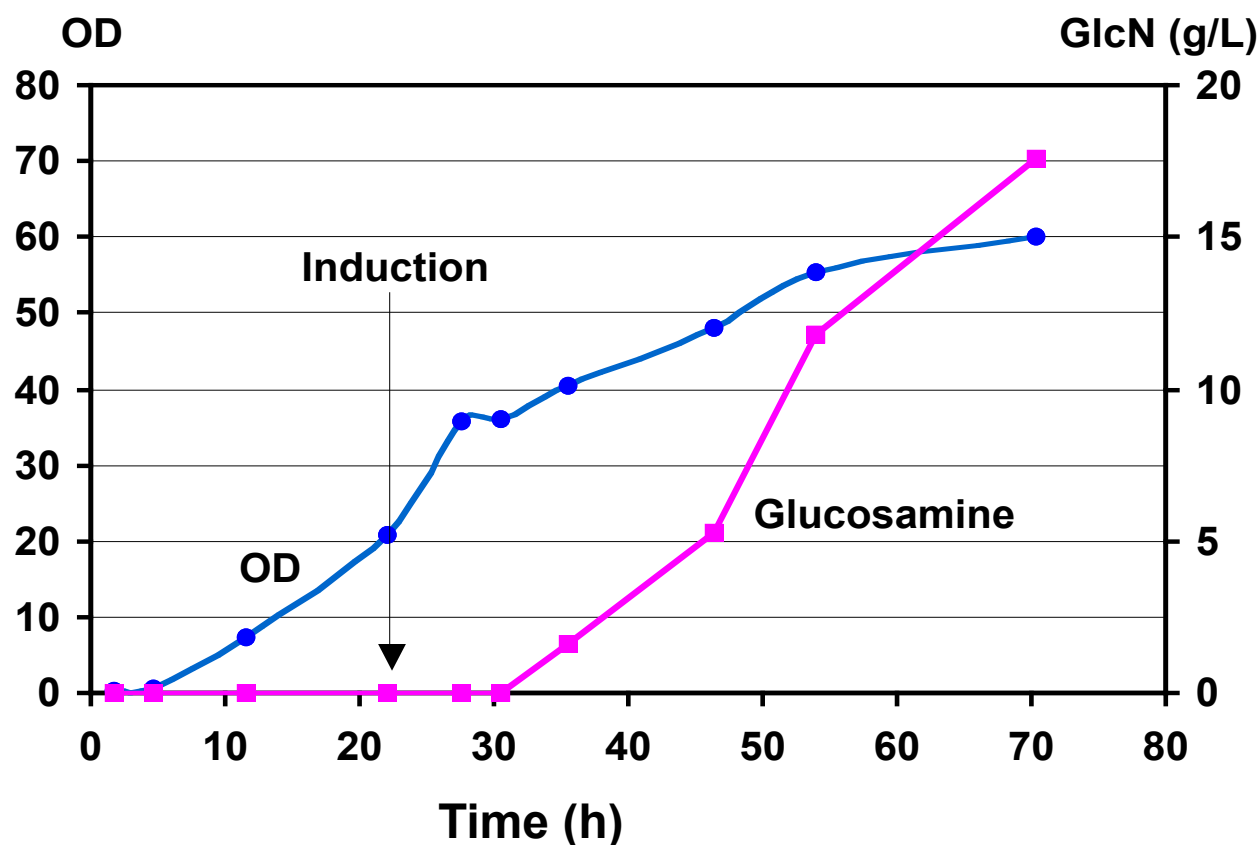
Locations of *glmS* Mutations and Invariant Residues



Mutations in the GImS Isomerase Domian



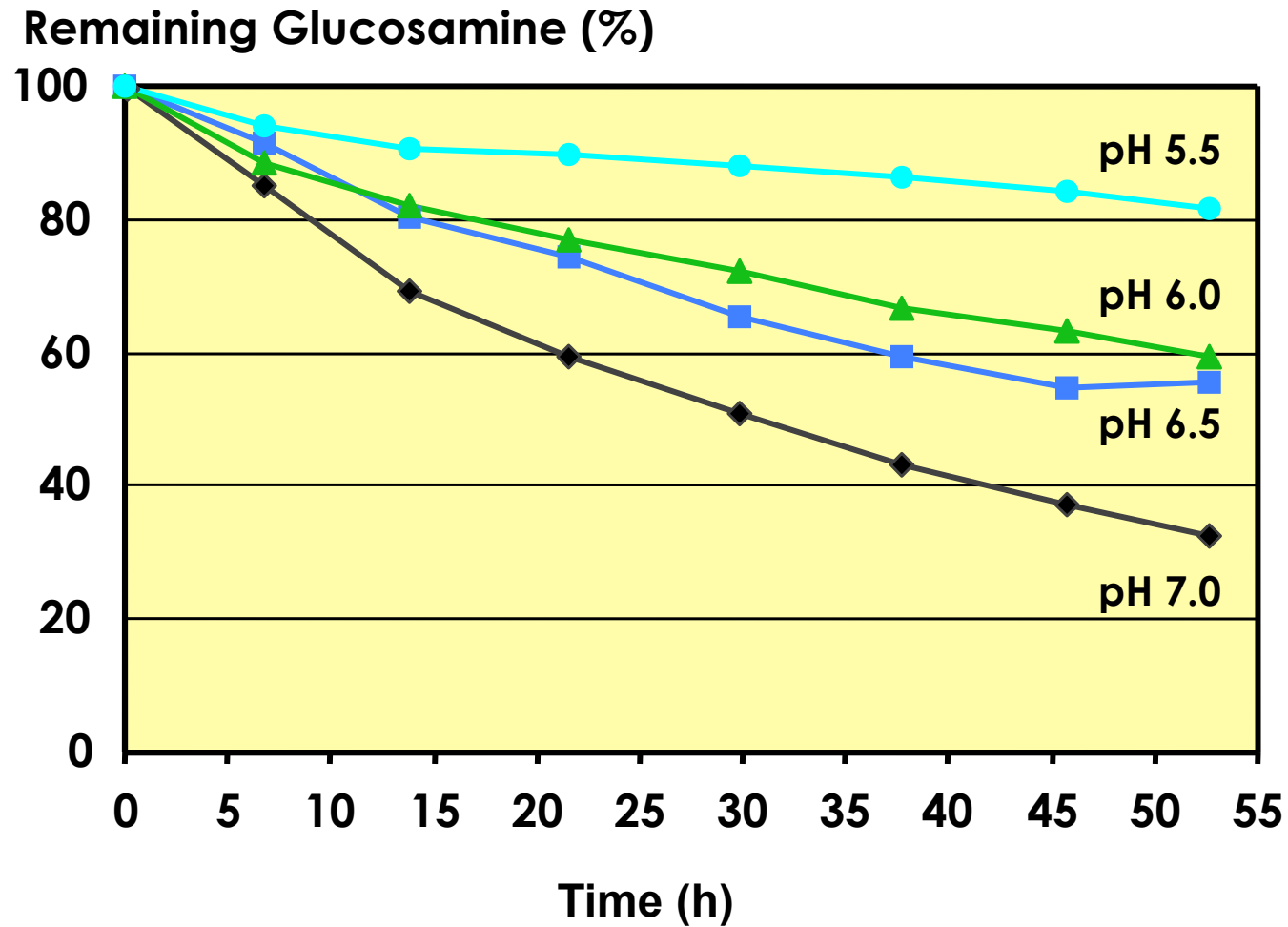
Lactose-Induced Glucosamine Production



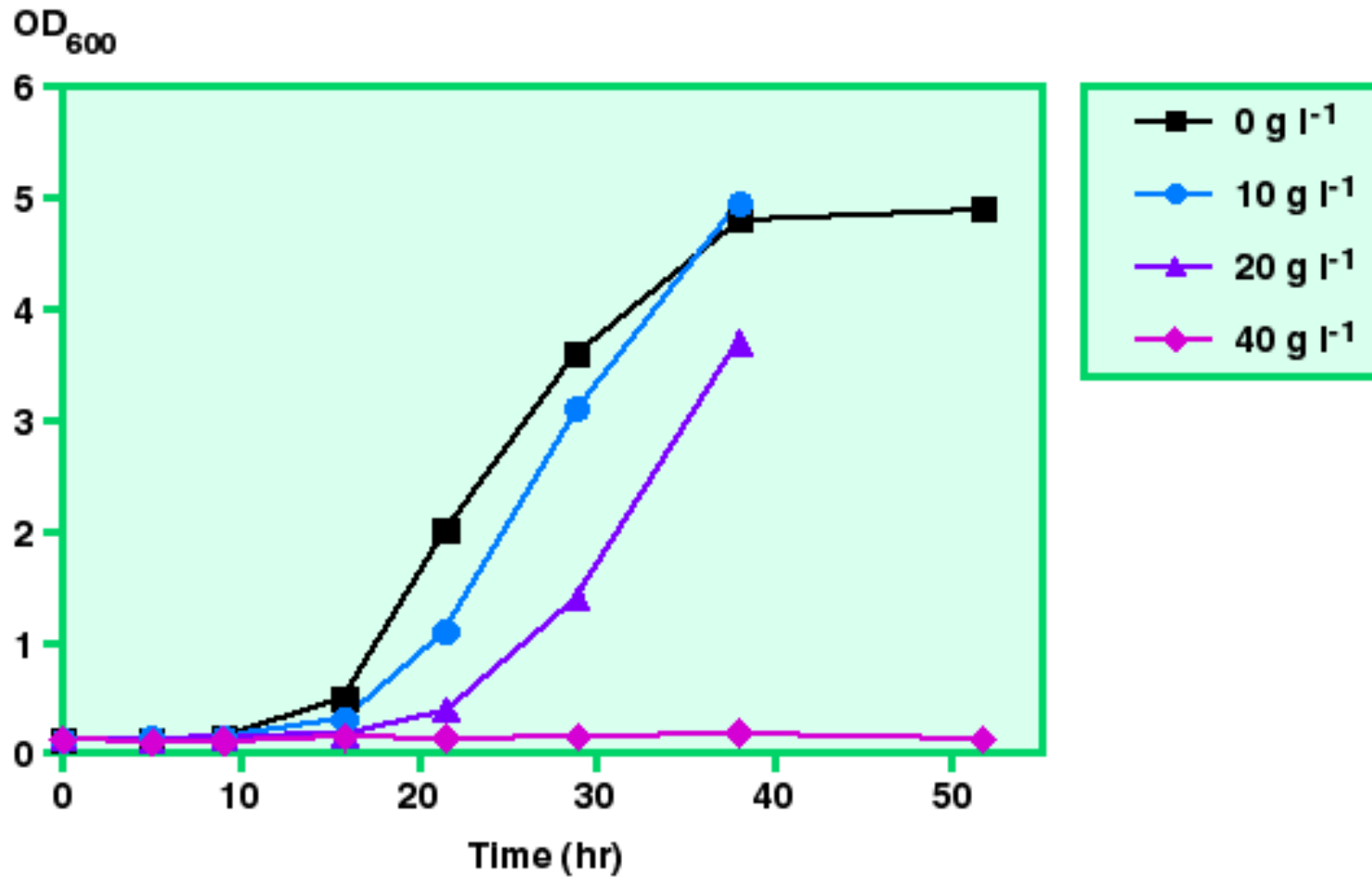
- Protocols for lactose induction of GlcN production were established in one-liter fermentors
- GlcN production reached 17 g/L, ~4,000-fold improvement over the wild type *E. coli* strain
- Titer was still far below the target for commercialization of 50 g/L



Glucosamine Degradation at Different pH

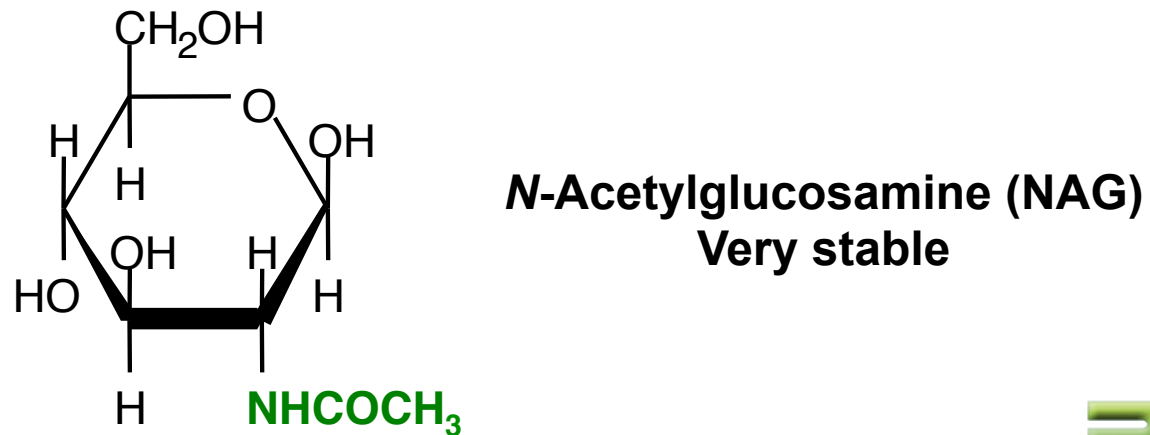
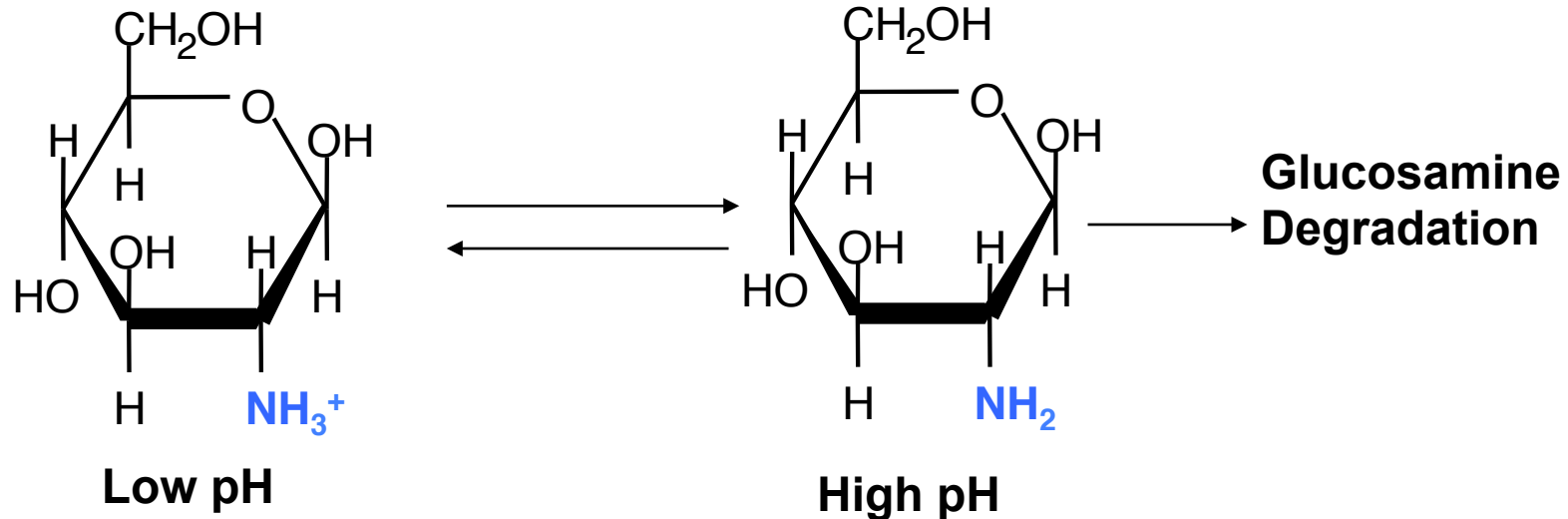


Growth Inhibition by Glucosamine at Different Concentrations



How to Overcome the Challenges?

Glucosamine Degradation and Toxicity

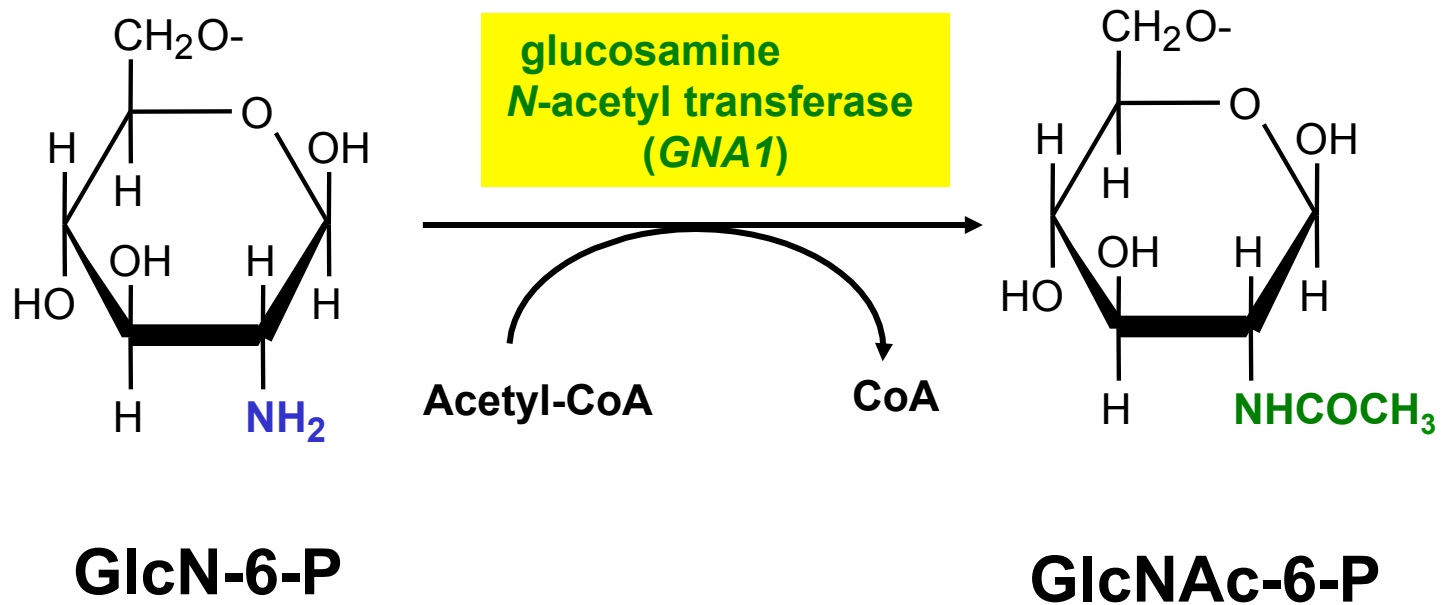


***N*-Acetylglucosamine as a Fermentation Product**

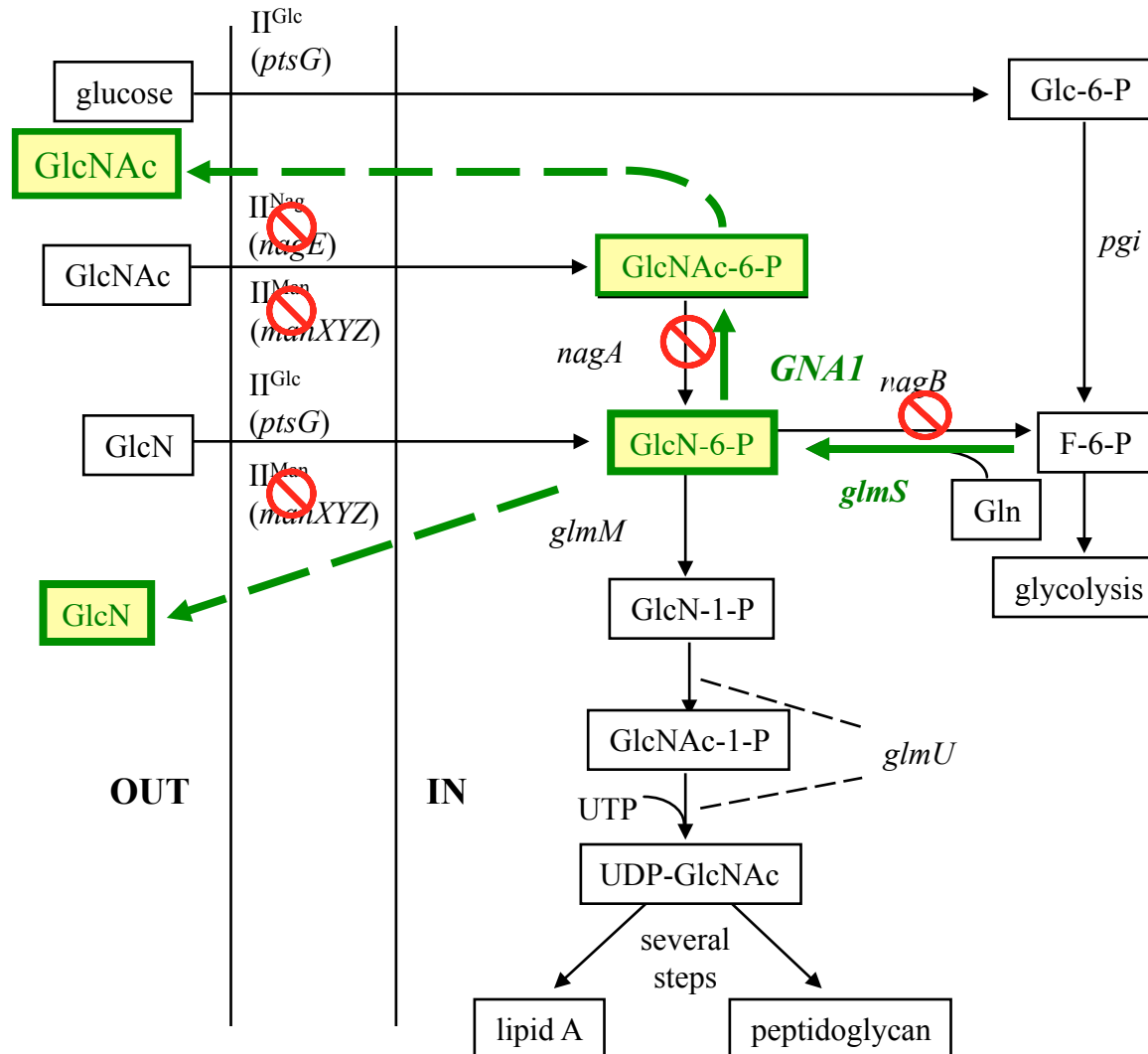
- ❖ **Favorable fermentation product characteristics**
 - Stable over a range of pH values
 - Non-inhibitory to growth
 - Readily converted to glucosamine by hydrolysis
- ❖ **Straightforward to modify the pathway**



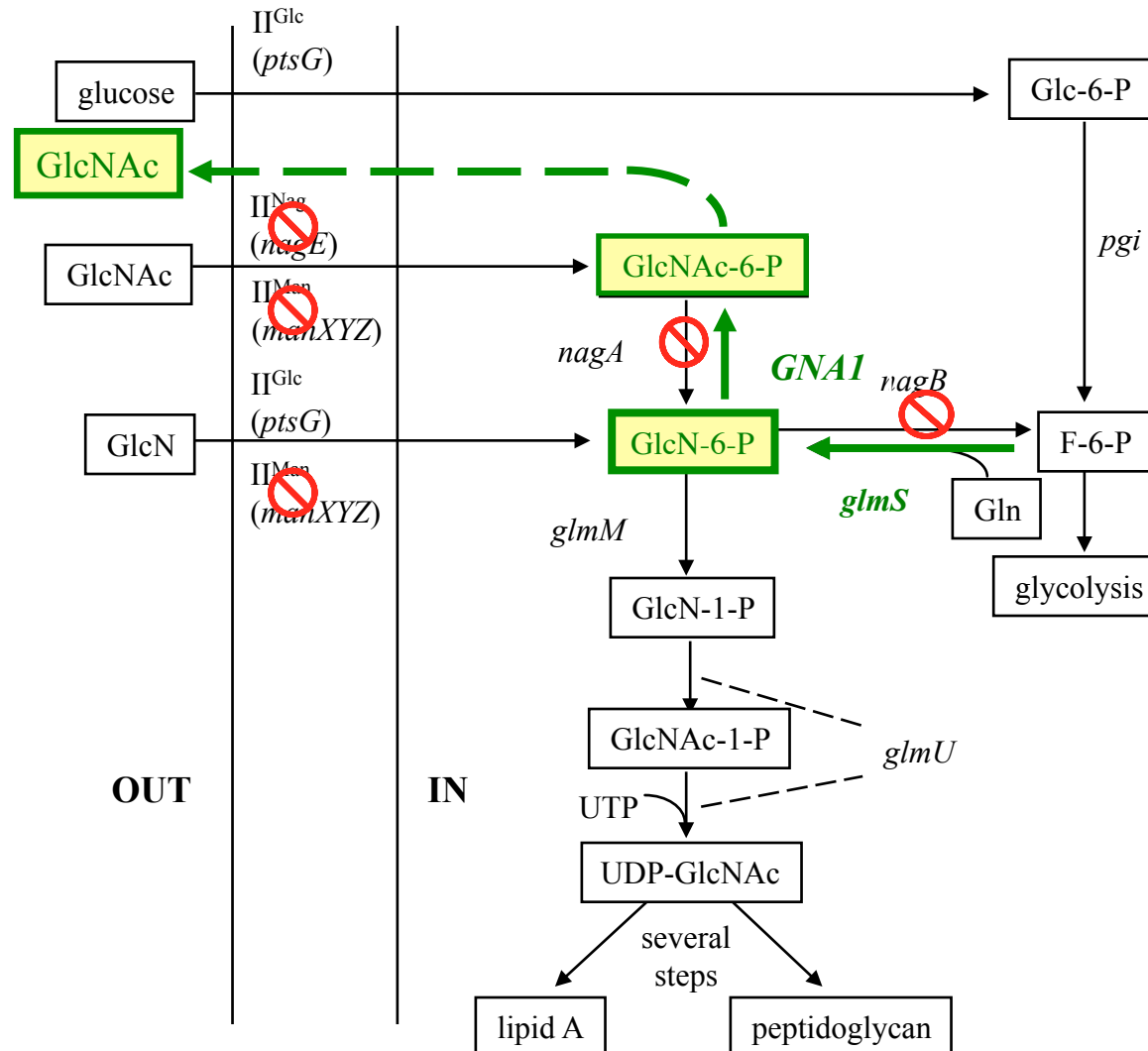
Over-Expression of a Yeast *GNA1* Gene



Metabolic Engineering to Produce *N*-Acetylglucosamine



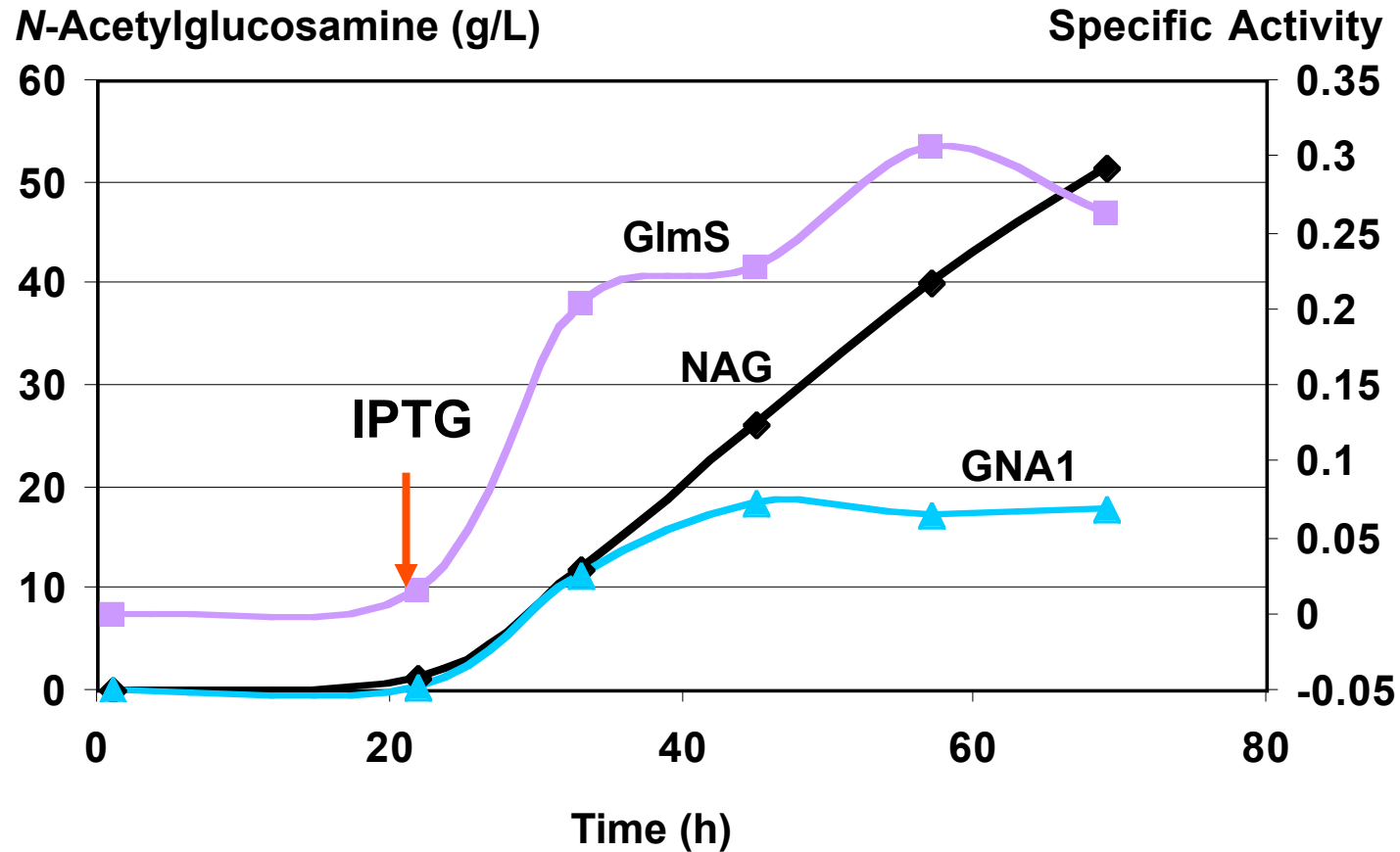
Metabolic Engineering to Produce *N*-Acetylglucosamine



- GlcNAc: accumulated in the medium as the only amino sugar product
- GlcN: not detectable

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NAG Fermentation with a GNA1 Plasmid



Process Parameters for *N*-Acetylglucosamine Fermentation

❖ Physical

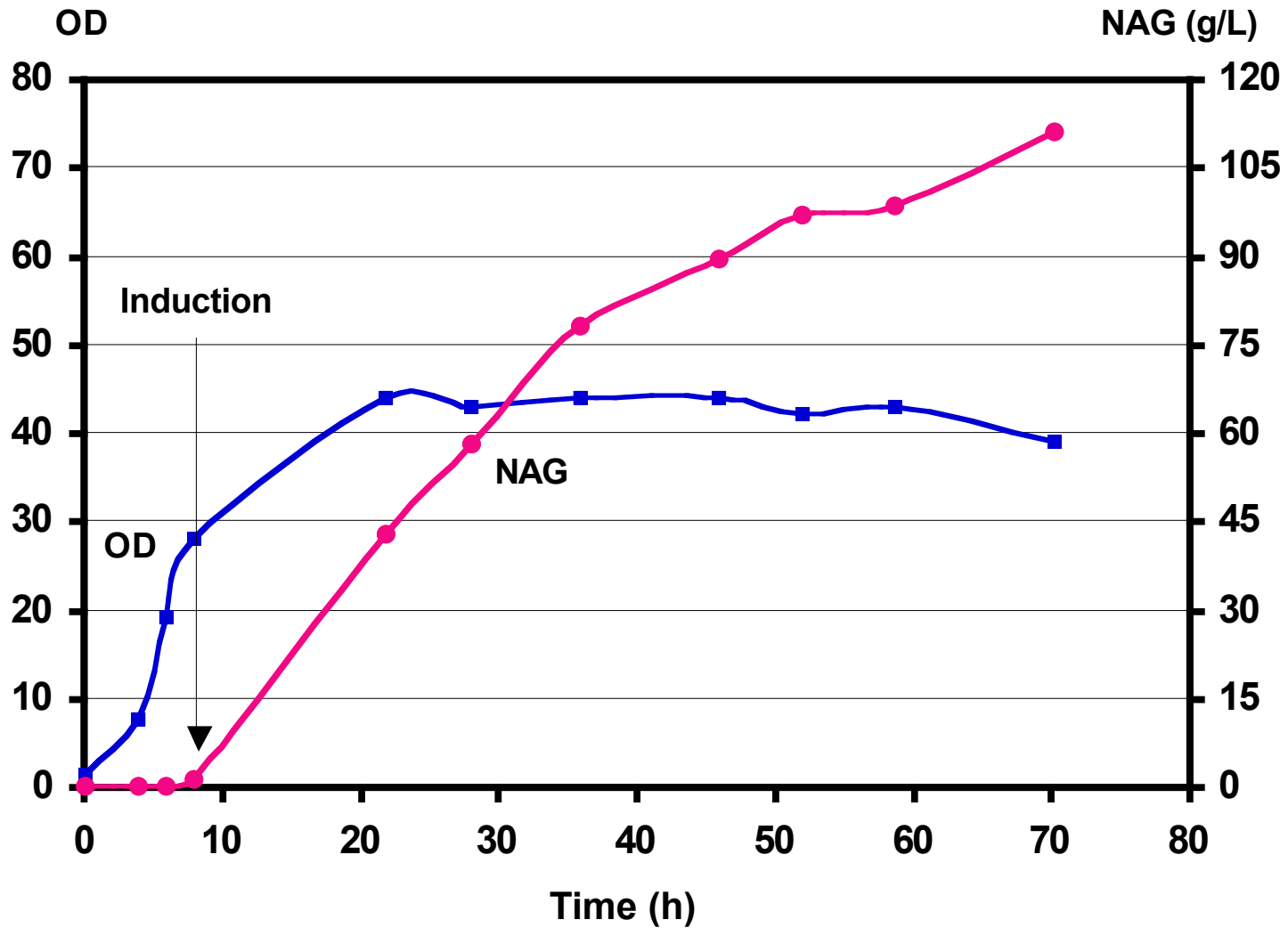
- Temperature: 30°C
- pH: 6.9 (controlled with ammonia)
- Oxygen: 20% or higher

❖ Process

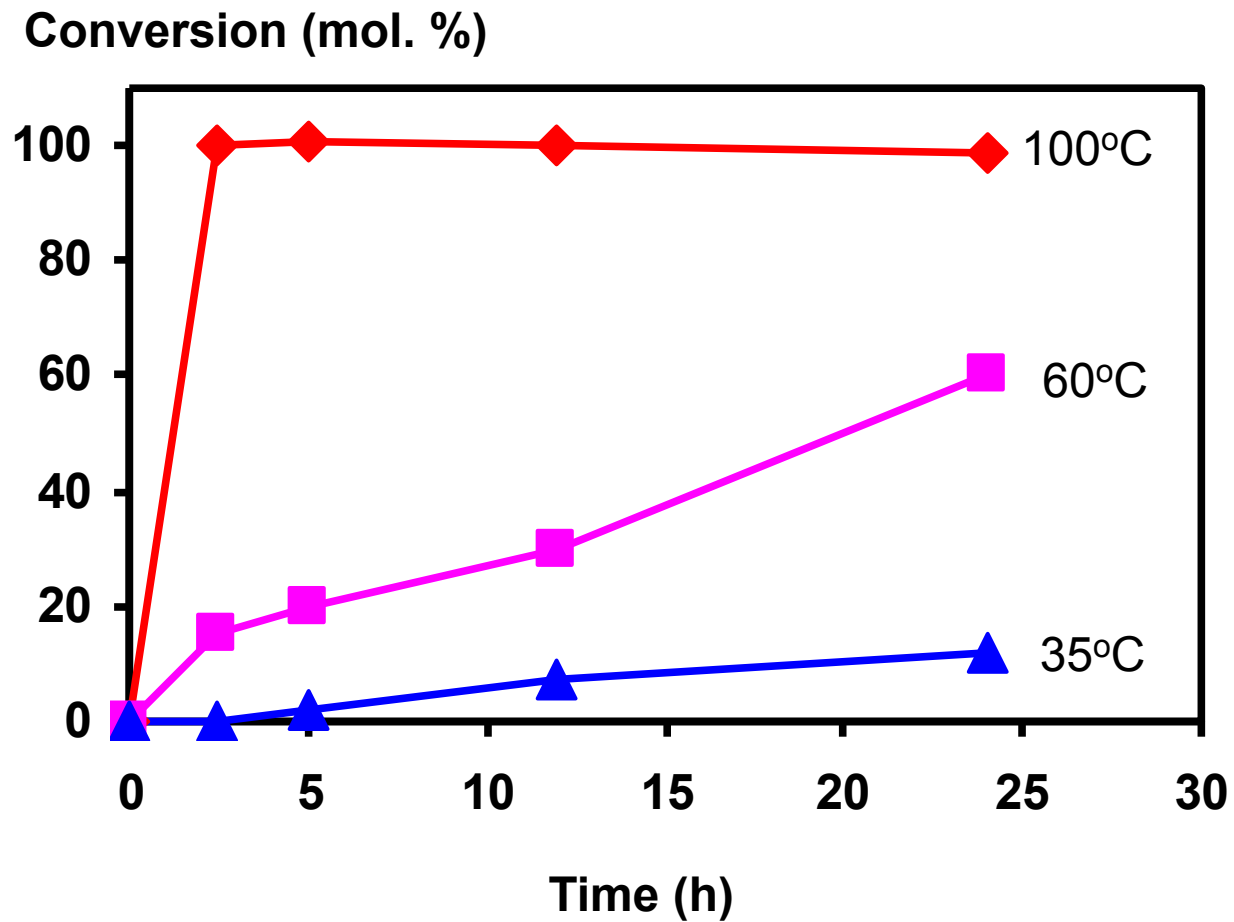
- Medium: Defined, minimal salts + glucose and NH₃
- Protocol: Bi-phasic (growth and production phases)
- Induction: Lactose
- Cell Density: 25 g/L



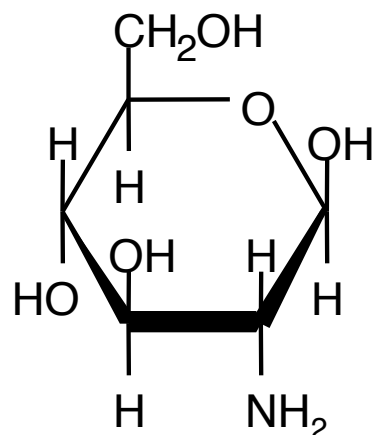
Lactose-Induced *N*-Acetylglucosamine Production



N-Acetylglucosamine Hydrolysis to Glucosamine



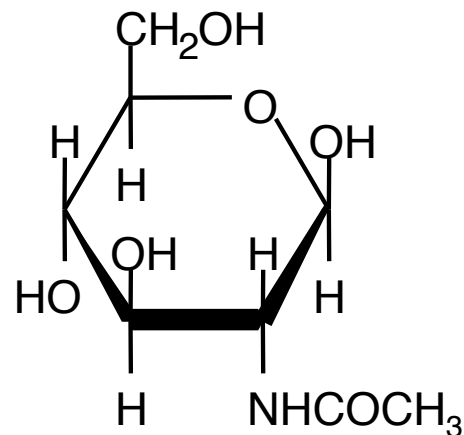
Glucosamine



- ❖ Increasingly used nutraceutical to treat arthritis
- ❖ Current manufacture by hydrolysis of chitin/chitosan from shellfish

- Raw material supply limitation and shellfish allergy problem
- Fermentation processes developed by Bio-Technical Resources

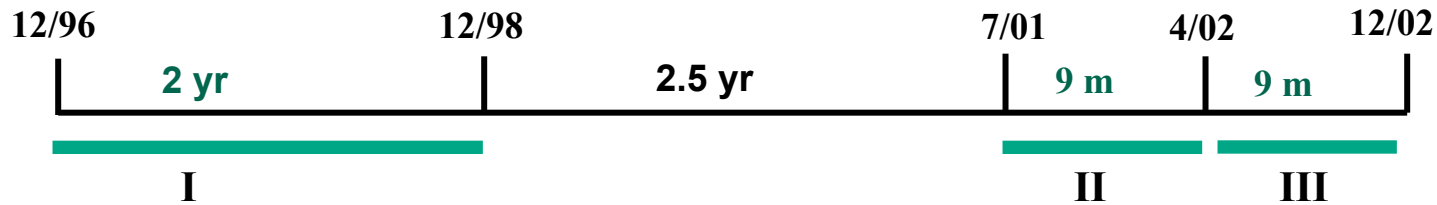
N-Acetylglucosamine



- ❖ Many nutraceutical, pharmaceutical and cosmetic applications
- ❖ Current manufacture by chemical *N*-acetylation of glucosamine



Technology Development Time Line



❖ Phase I

- Metabolic engineering feasibility
- Generation of improved GlnS enzyme

❖ Phase II:

- Glucosamine process improvement
- Identification of the key problem: glucosamine degradation
- Concept of NAG fermentation

❖ Phase III:

- Modification of the pathway for NAG production
- Development of a high-performance NAG process



Acknowledgments

Phase I:

Rich Burlingame, Alan Berry, Jim Millis, Kathy Nielsen, Chris Pynnonen, Bonnie Walsh, Cheryl Barrett, Fernando Sanchez-Riera

Phases II and III:

Ming-De Deng, Sarah Wassink, Candice Leanna, Kathy Nielsen, Al Grund, Jeff Running, Linsheng Song, Dave Severson, Brian Huckins, Bonnie Walsh, Troy Lutze, Reinhardt Rosson

References:

- ❖ Deng MD, Severson DK, Grund AD, Wassink SL, Burlingame RP, Berry A, Running JA, Kunesh CA, Song L, Jerrell TA and Rosson RA. Metabolic Engineering of *Escherichia coli* for industrial production of glucosamine and N-acetylglucosamine. *Metabolic Engineering* 7:201-214 (2007)
- ❖ Deng MD, Grund AD, Wassink SL, Peng SS, Nielsen KL, Huckins BD, Walsh BL and Burlingame RP. Directed evolution and characterization of *Escherichia coli* glucosamine synthase. *Biochimie*, 2006, 88:419-429

